

# Rosetta Basics: IO and Navigating Rosetta

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# Goals for this Talk

## 1. General Rosetta Concepts:

- How do I run basic Rosetta applications?
- Input/Output: file types, options, etc.

## 2. Learn where things are in Rosetta

- Your working directory is independent of these Rosetta directories
  - (AKA your data is stored outside of Rosetta)
- Highly encourage everybody to follow along from your command line
- Full path = path to file from home directory
  - ( $\sim$  = /usr/people/molgraph/)
- Relative path = path to file from current working directory

- Please stop me for questions!

# How do we get Rosetta?

- <https://www.rosettacommons.org/software/license-and-download>
- Weekly Releases: (e.g. “2016.37”)
  - Latest version of the code, released roughly every week
  - Every revision passes scientific performance tests
- Numbered Releases (e.g. “3.9”)
  - A weekly release that’s relabeled, released roughly every 6 months
- All tutorials use version 2017.36.59679 (May 2018)
- Links to documentation, forum and demos:
  - <https://www.rosettacommons.org/docs/latest/Home>
  - <https://www.rosettacommons.org/demos/latest/Home>

```
cd ~/rosetta_workshop/tutorials/short_talks/RosettaIO/
```

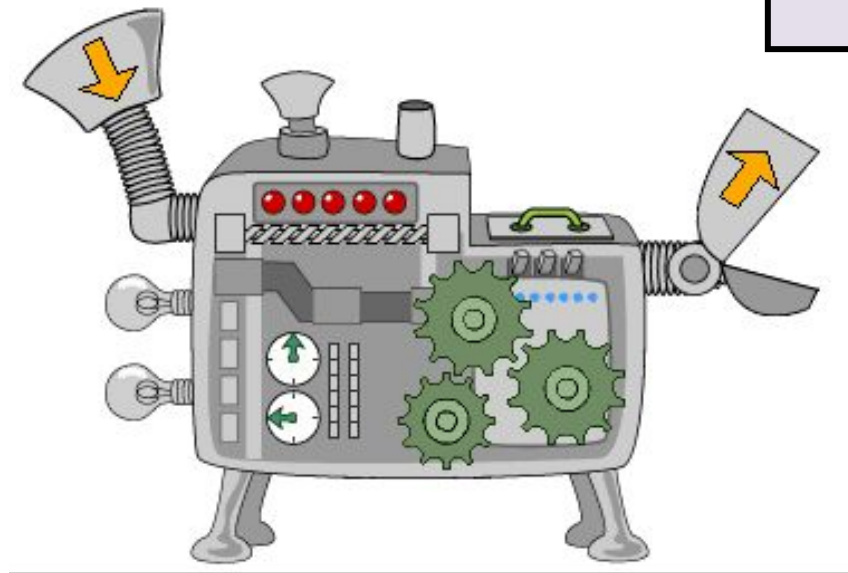
- Where you will find all the files associated with this talk
- input\_files/
- output\_files/
- scripts/
- Notice these files are located outside of Rosetta (aka you do NOT want to store your input/output files in directories within ~/rosetta\_workshop/rosetta/\*) !!

Tip: Open each file in your favorite text editor (gedit, vi, emacs, etc.) as we introduce them throughout the talk

# Basic Pipeline for Running a Rosetta Application

Input Files
Structure
Sequence
Application-Specific

Output Files
Structures
Scores
Log/Tracer



Rosetta Application

# Structure Files: PDB

Input Files
Structure
Sequence
Application-Specific

- PDBs are broken down by atom lines
- Each atom is characterized by the following (left to right across each row)
  - Atom, atom number, atom name, residue name, chain identifier, residue sequence number, x, y, z orthogonal coordinates, occupancy, temperature factor, element symbol, atom charge.
- Note: CIF not supported

## Snippet from 1qys.pdb

419	ATOM	57	N	ILE	A	10	3.073	-2.269	16.587	1.00	40.88	N
420	ATOM	58	CA	ILE	A	10	3.976	-3.337	16.187	1.00	40.88	C
421	ATOM	59	C	ILE	A	10	3.531	-4.697	16.730	1.00	40.88	C
422	ATOM	60	O	ILE	A	10	3.465	-4.900	17.944	1.00	70.73	O
423	ATOM	61	CB	ILE	A	10	5.399	-3.068	16.702	1.00	61.80	C
424	ATOM	62	CG1	ILE	A	10	5.870	-1.693	16.253	1.00	61.80	C
425	ATOM	63	CG2	ILE	A	10	6.343	-4.127	16.180	1.00	61.80	C
426	ATOM	64	CD1	ILE	A	10	7.241	-1.341	16.788	1.00	61.80	C
427	ATOM	65	N	ASP	A	11	3.225	-5.621	15.824	1.00	68.81	N
428	ATOM	66	CA	ASP	A	11	2.802	-6.960	16.220	1.00	68.81	C
429	ATOM	67	C	ASP	A	11	3.892	-7.970	15.878	1.00	68.81	C
430	ATOM	68	O	ASP	A	11	4.569	-7.850	14.850	1.00	68.81	O



## PDB files

- International standard
- Useful for a small number of structures
- One line per atom
- Readable by PyMol, MOE, Chimera, etc
- Must be processed with `clean_pdb.py` prior to use in Rosetta
- Full info: [www.wwpdb.org/documentation/file-formats](http://www.wwpdb.org/documentation/file-formats)

## Silent files

- Specific to Rosetta
- Useful for archiving many structures
- One line per residue
- Binary files: more compact, but not human-readable
- Silent --> PDB using `$ROSETTA/main/source/bin/extract_pdbs.linuxgccrelease`
- Full info: [https://www.rosettacommons.org/docs/latest/rosetta\\_basics/file\\_types/silent-file](https://www.rosettacommons.org/docs/latest/rosetta_basics/file_types/silent-file)



# Sequence Files:

- fasta: string of 1-letter amino acid names
- clustal: common for multiple sequence alignments
- grishin: RosettaCM-specific

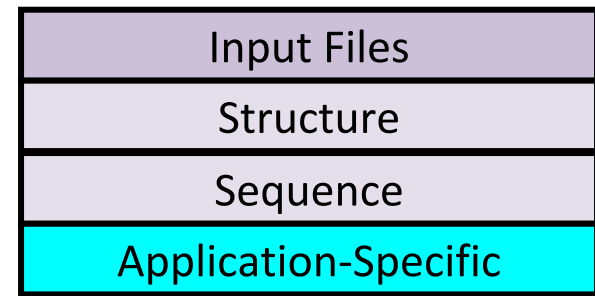
Input Files
Structure
Sequence
Application-Specific

1qys.fasta

```
>1QYS:A|PDBID|CHAIN|SEQUENCE
MGDIQVQVNIDDNGKNFDYTYTVTTESELQKVLNELMDYIKKQGAKRVRISITARTKKEA
EKFAAILIKVFAELGYNDIN
VTFDGDTVTVEGQLEGGSSLEHHHHHH
```

# Application-Specific Files:

- More details later during tutorials



- **Res Files: Indicates which residue positions should be designed**
- Params File: Custom parameters for small molecules or non-canonical amino acids
- Span File: Defines which residues are in the membrane
- Loops File: Identifies the loop residues for loop closure
- Constraint File: Experimentally derived restraints
- Fragment File: Short protein segments used for comparative modeling and de novo folding

# Now we have our input files...what do we do now?

```
$ROSETTA/main/source/bin/<your favorite Rosetta application> -option1  
<argument1> -option2 -option3 <argument3a> <argument3b>...
```

Note: Some options can take one or multiple arguments, while some don't take any.

# Now we have our input files...what do we do now?

```
$ROSETTA/main/source/bin/<your favorite Rosetta application> -option1  
<argument1> -option2 -option3 <argument3a> <argument3b>...
```

Note: Some options can take one or multiple arguments, while some don't take any.

```
>> $ROSETTA/main/source/bin/AbinitioRelax.linuxgccrelease  
-in:file:fasta ./input_files/1qys_A.fasta -in:file:native  
./input_files/1qys.pdb -in:file:frag3  
./input_files/aalelwA03_05.200_v1_3 -in:file:frag9  
./input_files/aalelwA09_05.200_v1_3 -abinitio:relax -relax:fast  
-abinitio::increase_cycles 10 -abinitio::rg_reweight 0.5  
-abinitio::rsd_wt_helix 0.5 -abinitio::rsd_wt_loop 0.5  
-use_filters true -psipred_ss2 ./input_files/1elwA.psipred_ss2  
-kill_hairpins -out:file:silent ./output_files/1qys_10.out  
-out:file:scorefile ./output_files/1qys_silent_scores.sc -nstruct  
10
```

# Now we have our input files...what do we do now?

```
$ROSETTA/main/source/bin/<your favorite Rosetta application> -option1  
<argument1> -option2 -option3 <argument3a> <argument3b>...
```

Note: Some options can take one or multiple arguments, while some don't take any.

```
>> $ROSETTA/main/source/bin/AbinitioRelax.linuxgccrelease  
-in:file:fasta ./input_files/1qys_A.fasta -in:file:native  
./input_files/1qys.pdb -in:file:frag3  
./input_files/aalelwA03_05.200_v1_3 -in:file:frag9  
./input_files/aalelwA09_05.200_v1_3 -abinitio:relax -relax:fast  
-abinitio::increase_cycles 10 -abinitio::rg_reweight 0.5  
-abinitio::rsd_wt_helix 0.5 -abinitio::rsd_wt_loop 0.5  
-use_filters true -psipred_ss2 ./input_files/1elwA.psipred_ss2  
-kill_hairpins -out:file:silent ./output_files/1qys_10.out  
-out:file:scorefile ./output_files/1qys_silent_scores.sc -nstruct  
10
```

## \$ROSETTA/main/source/bin/AbinitioRelax.linuxgccrelease @options.txt

- Contains all the same options as the command line, just in a more user-friendly fashion
- Easier to read, edit, log, etc.

### options.txt

```
## This is a comment
```

```
## input files
```

```
-in:file
```

```
-fasta ./input_files/1qys_A.fasta ## specifies location of fasta sequence file
```

```
-native ./input_files/1qys.pdb ## specifies location of native PDB; "native" files are often used to compare final output structures
```

```
-frag9 ./input_files/aa1elwA03_05.200_v1_9 ## not in this directory, but are 9-mer residue fragments generated on Robetta server
```

```
-frag3 ./input_files/aa1elwA03_05.200_v1_3 ## not in this directory, but are 3-mer residue fragments generated on Robetta server
```

```
-psipred_ss2 ./input_files/1elwA.psipred_ss2 ## not in this directory, but secondary structure prediction outputs from Robetta server
```

```
## application-specific flags
```

```
-abinitio:relax
```

```
-relax:fast
```

```
-abinitio::increase_cycles 10
```

```
-abinitio::rg_reweight 0.5
```

```
-abinitio::rsd_wt_helix 0.5
```

```
-abinitio::rsd_wt_loop 0.5
```

```
-use_filters true
```

```
-kill_hairpins
```

```
## output options
```

```
-out:file:silent ./output_files/1qys_10.out
```

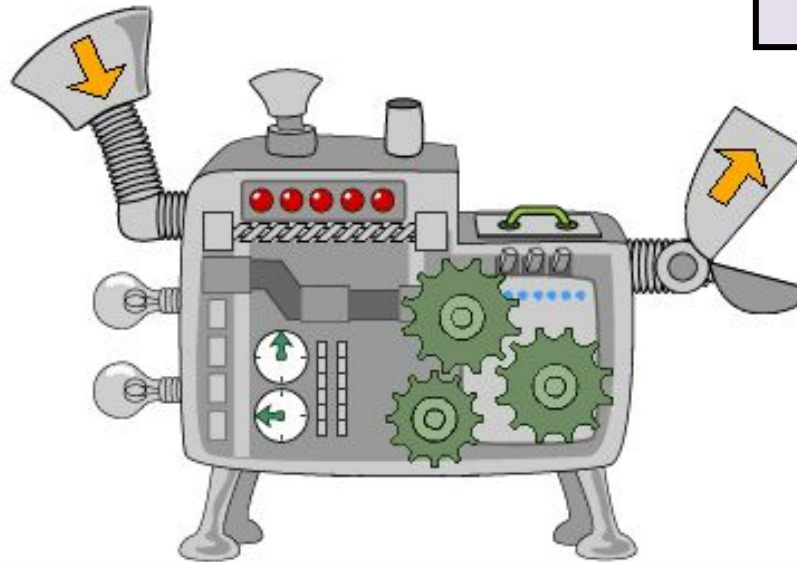
```
-out:file:scorefile ./output_files/1qys_silent_scores.sc
```

```
-nstruct 10
```

# Basic Pipeline for Running a Rosetta Application

Input Files
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```
$ROSETTA/main/source/bin/AbinitioRelax.linuxgccrelease @options.txt
```

Output Files

Scores

Structures

Log/Tracer

## Score Files:

- tab-delimited file containing all score terms for each output

./output\_files/1qys\_silent\_scores.sc

	SCORE:	score	fa_atr	fa_rep	fa_sol	fa_intra_rep	fa_elec	...	ref	time	description
1	SCORE:	-145.658	-416.906	48.038	235.048	1.023	-47.764	...	-12.643	121.000	1qys_0001
2	SCORE:	-145.660	-420.838	48.025	238.756	1.060	-46.720	....	-12.643	119.000	1qys_0002
3	SCORE:	-147.668	-414.585	46.523	235.308	1.033	-48.171	...	-12.643	115.000	1qys_0003
4	SCORE:	-147.324	-413.085	46.068	232.161	1.025	-46.810	...	-12.643	116.000	1qys_0004
5	SCORE:	-148.283	-423.099	47.967	241.560	1.046	-48.530	...	-12.643	110.000	1qys_0005
6	SCORE:	-147.020	-418.127	47.198	237.669	1.018	-48.364	...	-12.643	121.000	1qys_0006
7	SCORE:	-148.368	-417.523	46.620	235.293	1.049	-47.662	...	-12.643	111.000	1qys_0007
8	SCORE:	-147.644	-414.716	45.941	232.969	1.038	-46.471	...	-12.643	119.000	1qys_0008
9	SCORE:	-147.763	-416.130	46.386	235.877	1.023	-47.914	...	-12.643	118.000	1qys_0009
10	SCORE:	-146.560	-421.948	49.483	239.471	1.031	-49.106	...	-12.643	115.000	1qys_0010



Output Files

Scores

Structures

Log/Tracer

# Structure Files:

- PDB or silent file -- same as input files, but note that output PDBs contain per residue score terms after ATOM lines.

./output\_files/1qys\_0001.pdb

```
1509 #BEGIN_POSE_ENERGIES_TABLE 1qys_0001.pdb
1510 label fa_atr fa_rep fa_sol fa_intra_rep fa_elec pro_close hbond_sr_bb hbond_lr_bb hbond_bb_sc hbond_sc dslf_fa13 rama omega fa_dun p_aa_pp yhh_planarity ref linear_chainbreak overlap_chainbreak total
1511 weights 1 0.55 0.9375 0.005 0.875 1.25 1.17 1.17 1.17 1.1 1.25 0.25 0.625 0.7 0.4 0.625 1 1.33333 1 NA
1512 pose -423.638 49.1168 241.309 1.0398 -46.1456 0 -25.4907 -26.9979 -3.93408 -11.2344 0 -4.90455 4.21053 109.662 -13.6029 0.23024 -12.6427 0 0 -163.023
1513 ASP:NtermProteinFull_1 -2.66563 0.27015 2.41637 0.02503 -0.26885 0 0 0 0 -0.32408 0 0 0.0102 1.89419 0 0 -1.63002 0 0 -0.27263
1514 ILE_2 -5.61825 0.23689 2.80238 0.02592 -0.12286 0 0 0 0 0 -0.26155 0.00661 0.81411 -0.34843 0 1.0806 0 0 -1.38458
1515 GLN_3 -4.84167 0.48106 4.18521 0.00631 -1.21224 0 0 0 0 -0.48562 0 0.06079 0.00207 2.96363 0.1074 0 -1.51717 0 0 -0.25024
1516 VAL_4 -4.61075 0.23581 1.49989 0.01618 -0.54587 0 0 0 0 0 -0.24278 0.0034 0.03458 -0.42478 0 0.97964 0 0 -3.05468
1517 GLN_5 -4.16136 0.16768 3.36407 0.00633 -1.06934 0 0 0 0 -0.60901 0 0.04214 0.02118 2.77942 0.12364 0 -1.51717 0 0 -0.85242
1518 VAL_6 -5.59614 0.88153 1.82246 0.014 -0.69849 0 0 0 0 0 -0.2164 0.00792 0.02014 -0.46296 0 0.97964 0 0 -3.24828
1519 ASN_7 -4.85911 0.20247 3.95038 0.00414 -1.45872 0 0 0 0 0 -0.60901 0 -0.0151 0.00501 3.51664 0.05469 0 -1.19118 0 0 -0.39979
1520 ILE_8 -5.46549 0.36001 1.8693 0.02274 -0.68642 0 0 0 0 0 -0.22814 0.00246 0.20363 -0.48548 0 1.0806 0 0 -3.3268
1521 ASP_9 -3.90212 0.1626 3.5745 0.00384 -1.65188 0 0 0 0 -0.46353 0 0.12839 0.04956 2.18701 0.36117 0 -1.63002 0 0 -1.18048
1522 ASP_10 -3.84216 0.45653 3.22502 0.01376 -0.64368 0 0 0 0 -0.41813 0 0.55824 0.52659 1.7871 -0.93152 0 -1.63002 0 0 -0.89825
1523 ASN_11 -1.70761 0.57979 1.45451 0.00575 0.08728 0 0 0 0 0 0.70672 0.03429 1.84768 -0.73549 0 -1.19118 0 0 1.08173
1524 GLY_12 -0.80224 0.12458 0.5163 1e-05 0.04398 0 0 0 0 0 -0.35846 0.02351 0 -0.8684 0 0.17333 0 0 -1.1474
1525 LYS_13 -3.51045 0.33392 1.99225 0.00495 -0.05999 0 0 0 0 0 0.39242 0.00549 1.4125 -0.12885 0 -0.35857 0 0 0.08368
1526 ASN_14 -2.79919 0.07672 2.07492 0.00412 -0.26575 0 0 0 0 0 0.01568 0.00684 2.02524 0.20226 0 -1.19118 0 0 0.14967
1527 PHE_15 -5.49746 0.25492 2.43742 0.03039 -0.62276 0 0 0 0 0 -0.19658 0.06517 2.03196 -0.15724 0 0.61937 0 0 -1.03481
1528 ASP_16 -2.92446 0.08288 2.94318 0.02006 -0.24577 0 0 0 0 0 0.05442 0.00203 1.72485 -0.29358 0 -1.63002 0 0 -0.26641
1529 TYR_17 -5.88808 0.44904 2.60429 0.03033 -0.15087 0 0 0 0 0 -0.09114 0.03337 2.10883 -0.04977 0.00033 0.1625 0 0 -0.79117
1530 THR_18 -1.9813 0.05965 1.85734 0.00877 -0.21462 0 0 0 0 0 0.02411 0.047 0.04309 -0.12051 0 0.20134 0 0 -0.07513
1531 TYR_19 -6.68966 0.80179 3.35153 0.0269 -1.08788 0 0 0 0 -0.41091 0 -0.20557 0.01282 1.67279 -0.30295 0.00237 0.1625 0 0 -2.66626
1532 THR_20 -2.62243 0.31036 2.76908 0.00722 -0.10022 0 0 0 0 -0.16154 0 0.12349 0.04786 0.09622 -0.08035 0 0.20134 0 0 0.59103
1533 VAL_21 -3.80377 0.75681 1.67726 0.01702 -0.122 0 0 0 0 0 -0.19285 0.00444 0.02274 -0.31811 0 0.97964 0 0 -0.97881
```

# Log Files/Tracer:

- Printed version of what runs on the terminal
- Keeps track of what's occurring during a simulation--  
VERY important for keeping records
- These files get large:
  - Silence certain tracers:
    - -mute core.chemical.ResidueTypeSet
  - Change verbosity level  
(Error/Warning/Info/Debug/Trace)
    - -out:levels all:Warning core.init:Info

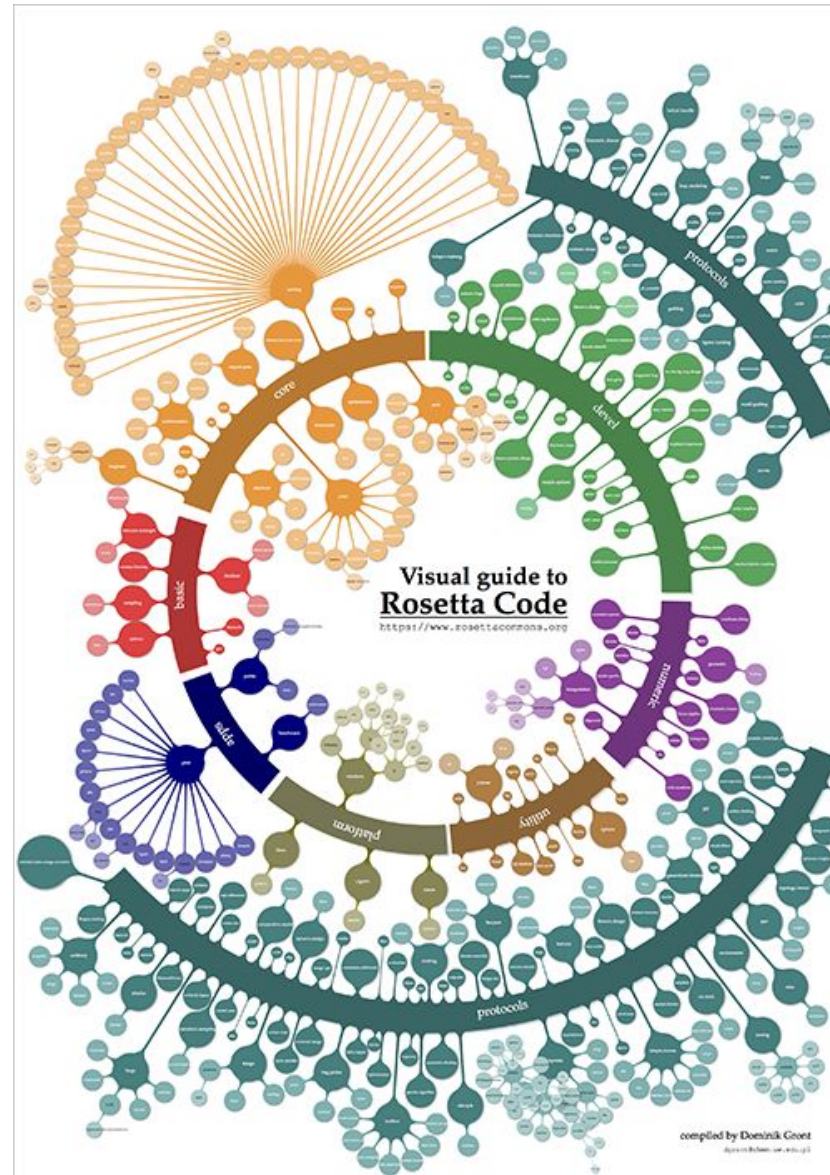
Output Files
Scores
Structures
Log/Tracer

`$ROSETTA/main/source/bin/AbinitioRelax.linuxgccrelease @options.txt >> run.log`

```
core.init: Rosetta version exported from http://www.rosettacommons.org
core.init: command: /dors/meilerlab/apps/rosetta/rosetta_2016.08.58479/main/source/bin/rosetta_scripts.default.linuxgccrelease @docking.options -parser:protocol docking
core.init: 'RNG device' seed mode, using '/dev/urandom', seed=1059677151 seed_offset=0 real_seed=1059677151
core.init.random: RandomGenerator:init: Normal mode, seed=1059677151 RG_type=mt19937
core.init: Resolved executable path: /dors/meilerlab/apps/rosetta/rosetta_2016.08.58479/main/source/build/src/release/linux/2.6/64/x86/gcc/5.2/default/rosetta_scripts.d
core.init: Looking for database based on location of executable: /dors/meilerlab/apps/rosetta/rosetta_2016.08.58479/main/database/
protocols.jd2.PDBJobInputter: Instantiate PDBJobInputter
protocols.jd2.PDBJobInputter: PDBJobInputter::fill_jobs
protocols.jd2.PDBJobInputter: pushed 3gbm_HA_3gbn_Ab.pdb nstruct indices 1 - 50
protocols.evaluation.ChiWellRmsdEvaluatorCreator: Evaluation Creator active ...
protocols.jd2.JobDistributor: Parser is present. Input mover will be overwritten with whatever the parser creates.
protocols.jd2.PDBJobInputter: PDBJobInputter::pose_from_job
protocols.jd2.PDBJobInputter: filling pose from PDB 3gbm_HA_3gbn_Ab.pdb
core.chemical.ResidueTypeSet: Finished initializing fa_standard residue type set. Created 384 residue types
core.chemical.ResidueTypeSet: Total time to initialize 0.43 seconds.
core.conformation.Conformation: Found disulfide between residues 7 461
...
protocols.rosetta_scripts.ParsedProtocol.REPORT: =====End report for =====
protocols.rosetta_scripts.ParsedProtocol.REPORT: =====Begin report for =====
protocols.rosetta_scripts.ParsedProtocol.REPORT: =====End report for =====
protocols.jd2.JobDistributor: 3gbm_HA_3gbn_Ab_full_0050 reported success in 381 seconds
protocols.jd2.JobDistributor: no more batches to process...
protocols.jd2.JobDistributor: 50 jobs considered, 50 jobs attempted in 16297 seconds
```

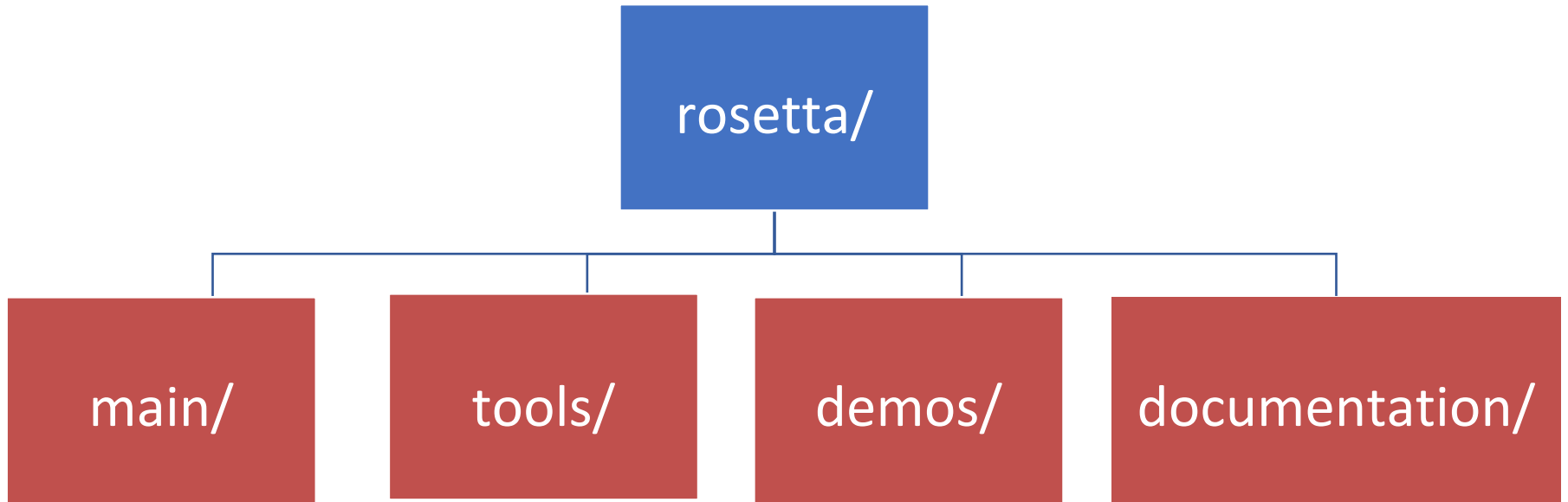
Any Questions?

# With great power comes great responsibility



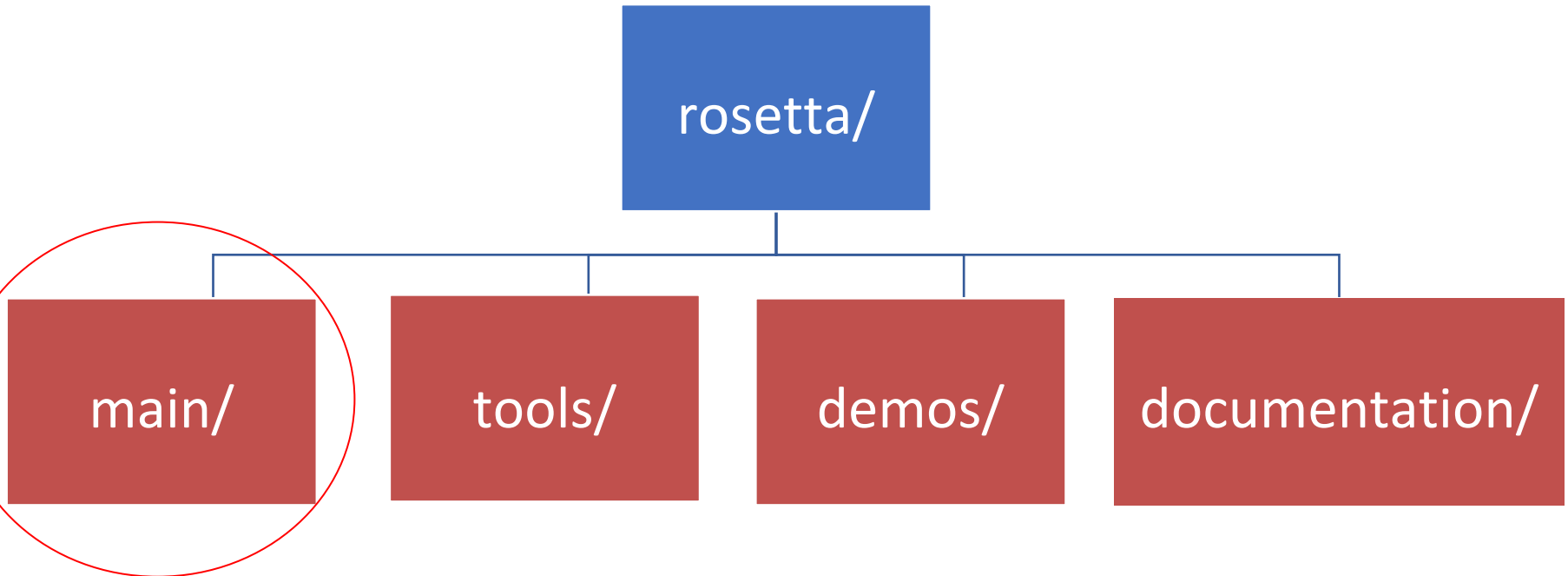
```
cd ~/rosetta_workshop/rosetta/
```

# Basic Rosetta Structure

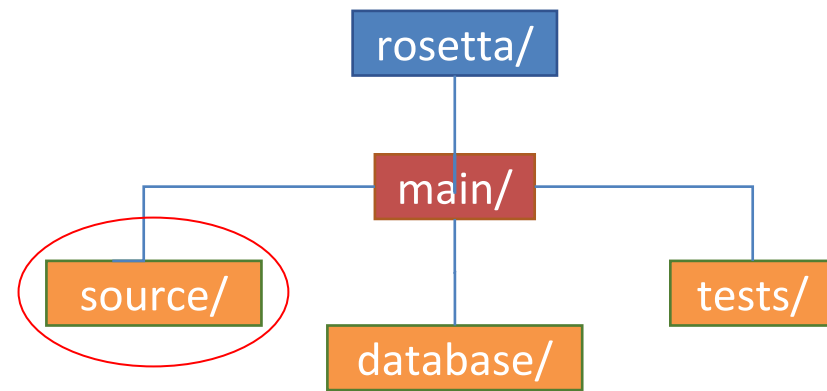


```
cd ~/rosetta_workshop/rosetta/
```

# Basic Rosetta Structure

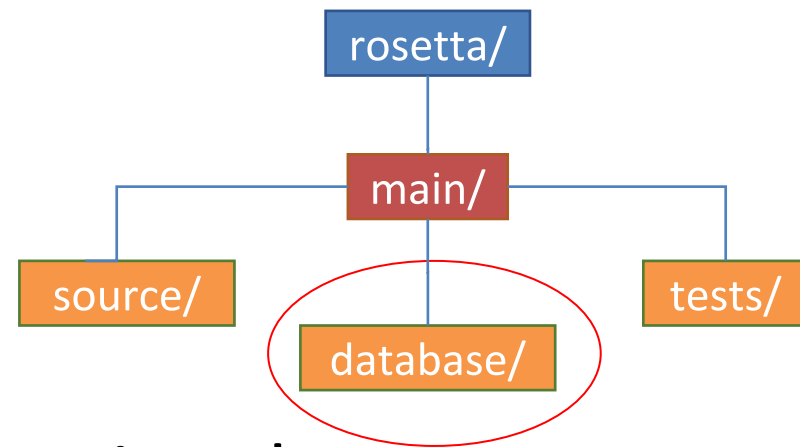


```
cd ~/rosetta_workshop/rosetta/main/source/
```



- **Rosetta/main/source/bin/**
  - Most applications you will run are calling programs within the bin directory
    - `rosetta_scripts.default.linuxgccrelease`
    - `score_jd2.default.linuxgccrelease`
    - `relax.default.linuxgccrelease`
- **Rosetta/main/source/scons.py**
  - Used for compiling
- **Rosetta/main/source/src/**
  - This is where all of the code lives

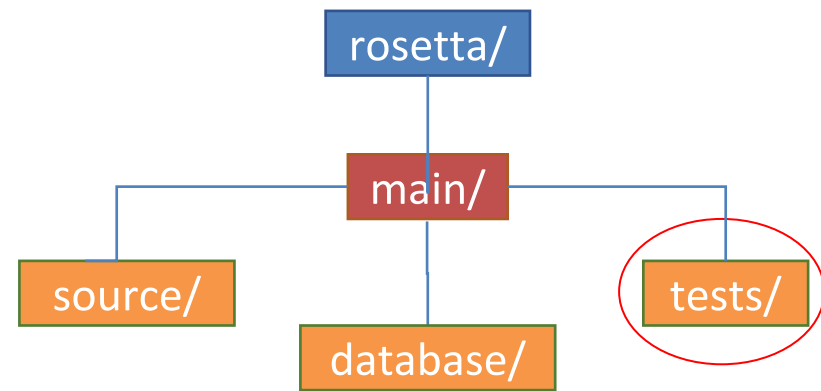
```
cd ~/rosetta_workshop/rosetta/main/database/  
(relative path from source/ == cd ../database/)
```



- Contains pre-defined information that Rosetta needs for runs
- [Rosetta/main/database/chemical/](#)
  - Residue information--params files
  - Atom sets
- [Rosetta/main/database/scoring/](#)
  - Default weights files
  - Rotamer libraries

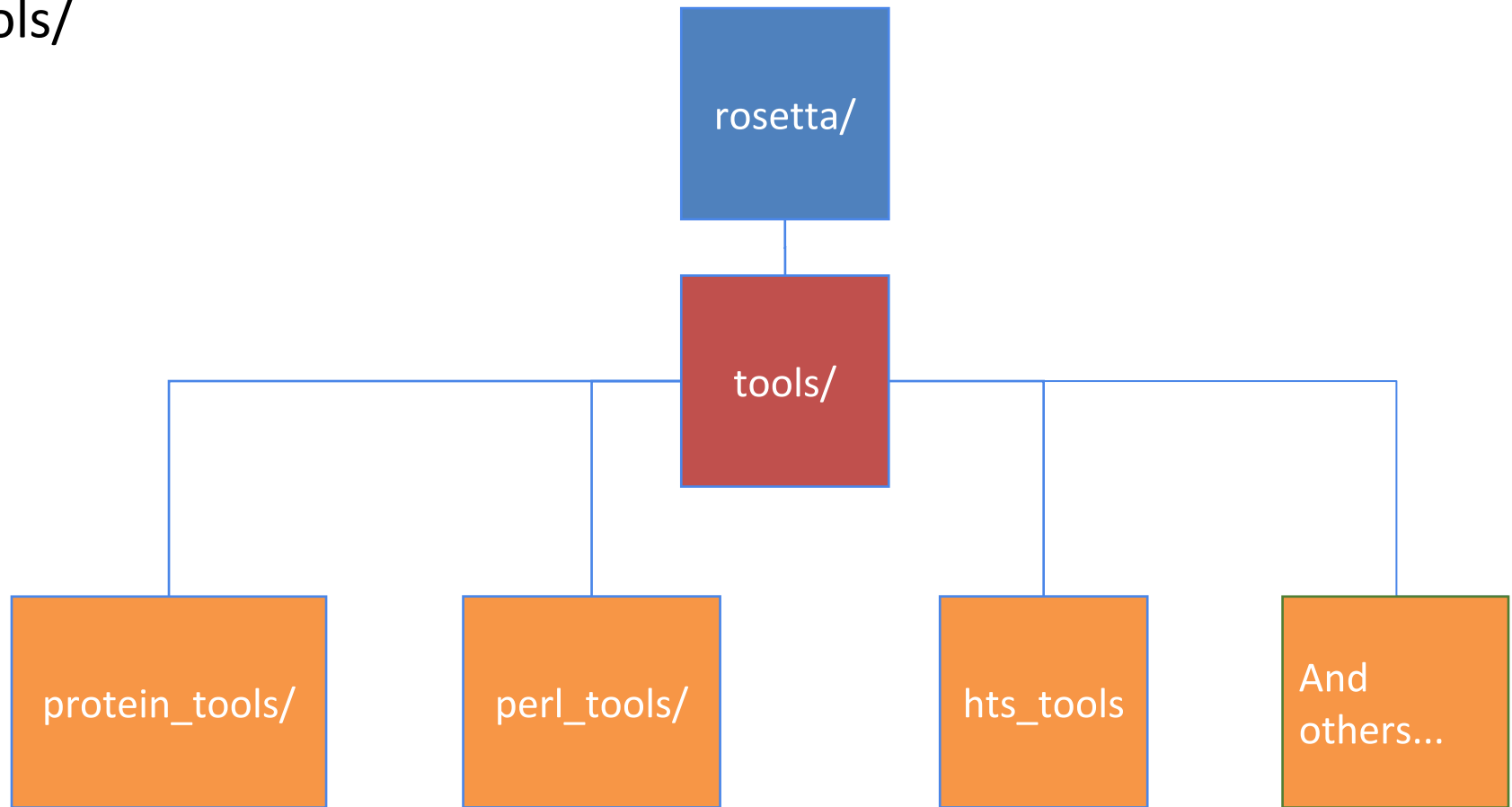


```
cd  
~/rosetta_workshop/rosetta/main/t  
ests/
```

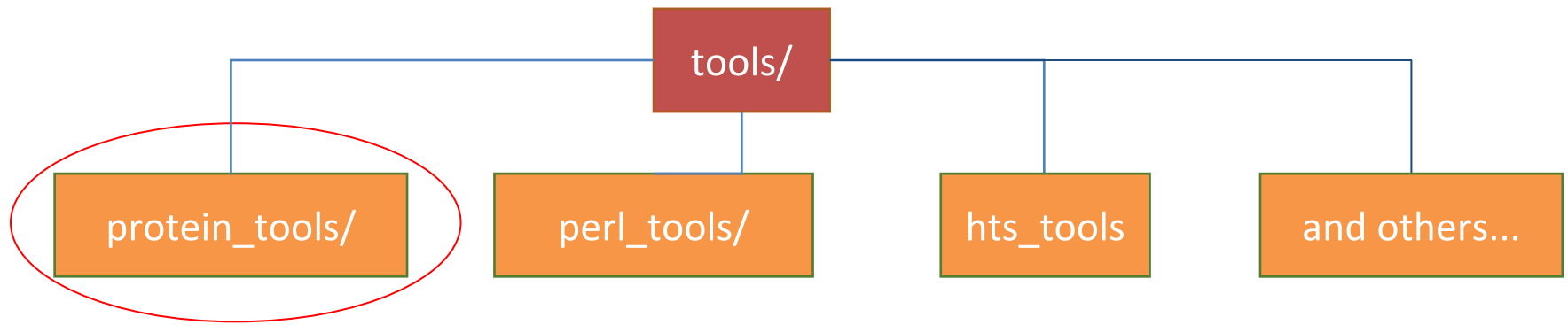


- [Rosetta/main/tests/](#)
  - This is for developer use--don't worry about this (fun to read though!)

```
cd  
~/rosetta_workshop/rosetta  
/tools/
```

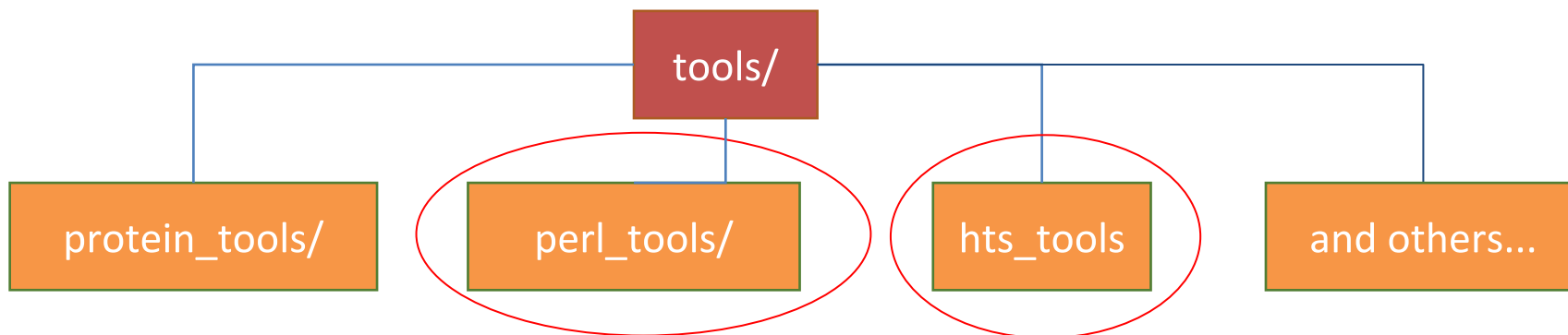


- These scripts are incredibly help for smaller, more basic tasks
- Used mainly to setup or analyze runs



## Rosetta/tools/protein\_tools/scripts/

- **clean\_pdb.py**
  - Makes a PDB “Rosetta-proof” and used at the beginning of almost any protocol
- `pdb_renumber.py`
- `score_vs_rmsd.py`
  - Setup for score vs. RMSD plots
- `top_n_percent.py`
  - Extracts tags (protein names) for top given percent of models based on score term



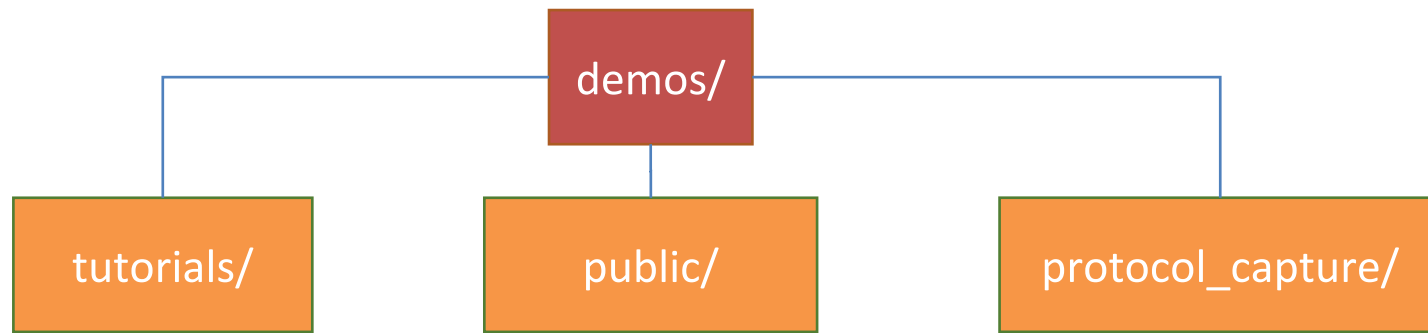
- **Rosetta/tools/perl\_tools/**

- addchain.pl
- getFastaFromCoords.pl
- getCAcoords.pl

- **Rosetta/tools/hts\_tools/**

Scripts used mainly to setup high throughput screen runs (> 1000 compounds)

- make\_params.py
- get\_descriptor\_data.py
- setup\_screening\_project.py

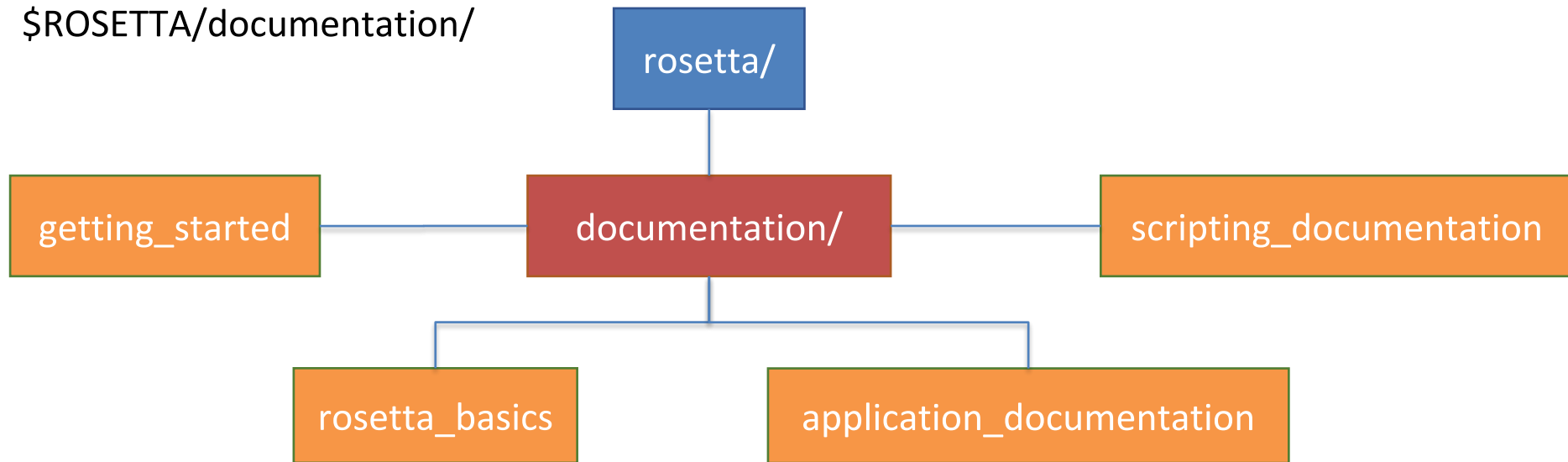


Input files/options are used in protocols--good place to start when you're doing learning a new method

- [Rosetta/demos/tutorials/](#)
  - these tutorials
- [Rosetta/demos/public/](#)
  - used to go through sample procedures
- [Rosetta/demos/protocol\\_capture/](#)
  - protocols associated with a publication

**DISCLAIMER: May be out of date, always check Wiki/RosettaCommons/forum for latest information!**

\$ROSETTA/documentation/



(Everything here is also on the RosettaCommons site!)

- Understanding general Rosetta concepts
  - Where to find FAQs (How long does this run take?)
  - Options list, file types
  - Protocols you can use
- General structural biology FAQs
  - How do I do X?

# Other Rosetta Resources:

- <https://www.rosettacommons.org/docs/latest/Home>
  - Forum
  - Documentation
  - User guides
  - Everything Rosetta-related you could ever dream of!

GO HAVE FUN!