Rosetta Basics: IO and Navigating Rosetta

Shannon Smith

shannon.t.smith.1@vanderbilt.edu

Graduate Student

Meiler Lab, Vanderbilt University

VANDERBILT **V** UNIVERSITY

Center for Structural Biology and Institute of Chemical Biology
Departments of Chemistry, Pharmacology, and Biomedical Informatics

Goals for this Talk

1. General Rosetta Concepts:
   - How do I run basic Rosetta applications?
   - Input/Output: file types, options, etc.

2. Learn where things are in Rosetta
   - Your working directory is independent of these Rosetta directories (AKA your data is stored outside of Rosetta)

QUESTIONS ARE ENCOURAGED!

Files for this talk are in ~/rosetta_workshop/tutorials/ short_talks/RosettaIO/

Notice these files are located outside of Rosetta (aka you do NOT want to store your input/output files in directories within ~/rosetta_workshop/rosetta/*) !!
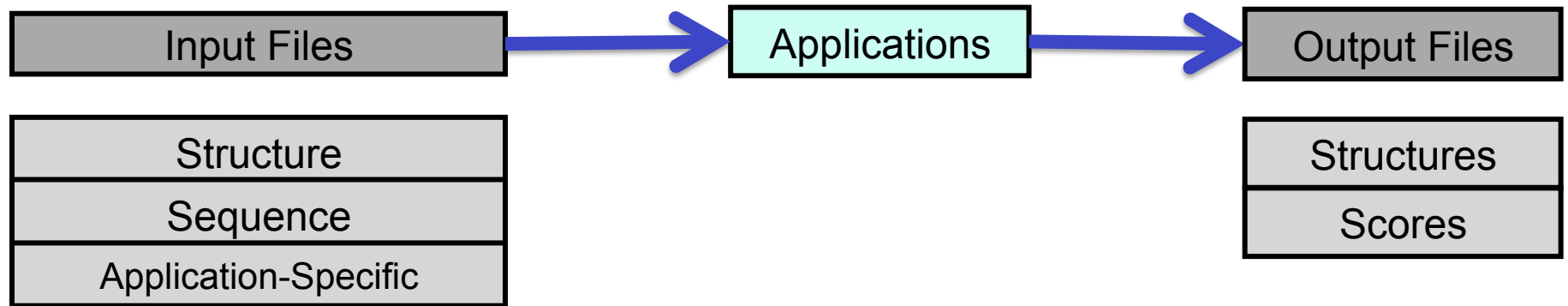
Tip: Open each file in your favorite text editor (gedit, vi, emacs, etc.) as we introduce them through the talk

How do I get Rosetta?

- https://www.rosettacommons.org/software/license-and-download

- Weekly Releases: (e.g. "2019.07")

  - Latest version of the code, released roughly every week

  - Every revision passes scientific performance tests

- Numbered Releases (e.g. "3.10")

  - A weekly release that's relabeled, released roughly every 6 months

- All tutorials use version 2019.07.60616

- Links to documentation, forum and demos:

  - https://www.rosettacommons.org/docs/latest/Home

  - https://www.rosettacommons.org/demos/latest/Home

General Workflow

```
┌─────────────────┐      ┌──────────────┐      ┌──────────────┐
│   Input Files   │ ───► │ Applications │ ───► │ Output Files │
└─────────────────┘      └──────────────┘      └──────────────┘

┌─────────────────┐                            ┌──────────────┐
│    Structure    │                            │  Structures  │
├─────────────────┤                            ├──────────────┤
│    Sequence     │                            │    Scores    │
├─────────────────┤                            └──────────────┘
│Application-Specific│
└─────────────────┘
```

Reading structures into Rosetta

PDB files

- International standard
- Readable by PyMol, MOE, Chimera, etc
- One line per atom
- Useful for small number of structures

Silent files

- Specific to Rosetta
- Compact
- One line per residue
- Useful for archiving many structures
- Binary silent files: more compact, but not human-readable

How do I run a Rosetta command?

Every command has the same basic layout:

Path to the application   Arguments

`<path_to_rosetta>/main/source/bin/<app_name>.default.linuxgccrelease` `-arg1 -arg2`

Example for you first Rosetta command:

`<path_to_rosetta>/main/source/bin/score_jd2.default.linuxgccrelease -in:file:s \`
`input_files/test1.pdb -out:file:scorefile output_files/test1.sc -out:pdb \`
`output_files/test1_out.pdb`

**Note: the "\" represents a continuous line

Common command line arguments

- Input options:
  - ➢ -parser:protocol example.xml ## RosettaScripts XML file
  - ➢ -in:file:s example_in.pdb ## input a PDB structure file
  - ➢ -in:file:fasta example.fasta ## input a FASTA sequence file
  - ➢ -in:file:silent example_in.silent ## input a Rosetta silent file
  - ➢ -in:file:extra_res_fa example.params ## input a parameters file for uncommon compounds
  - ➢ -nstruct 42 ## produce 42 outputs

- Output options:
  - ➢ -out:file:silent example_out.silent ## output structures to silent file
  - ➢ -out:file:scorefile example_out.sc ## output scorefile for run

Scores can be found in output scorefiles and PDBs

output_files/test1.sc

```
SEQUENCE:
SCORE:     score     fa_atr     fa_rep     fa_sol   fa_intra_rep   fa_elec   ...    omega    fa_dun     p_aa_pp       ref              description
SCORE:  -1217.209  -2778.696   266.309   1545.149        5.900   -301.320   ...   63.032   684.989   -109.110   -32.534   3gbm_HA_3gbn_Ab_full_0011
SCORE:  -1217.028  -2792.422   263.906   1549.738        5.867   -295.799   ...   66.036   682.694   -108.402   -32.534   3gbm_HA_3gbn_Ab_full_0012
SCORE:  -1204.280  -2760.354   259.175   1534.072        5.913   -293.050   ...   65.391   674.840   -108.393   -32.534   3gbm_HA_3gbn_Ab_full_0013
SCORE:  -1207.127  -2768.191   260.443   1541.857        5.881   -301.847   ...   67.951   686.381   -110.919   -32.534   3gbm_HA_3gbn_Ab_full_0014
SCORE:  -1208.390  -2769.872   262.398   1539.668        5.879   -297.571   ...   64.073   681.731   -109.633   -32.534   3gbm_HA_3gbn_Ab_full_0015
```

Bottom of output_files/test1.pdb

```
ATOM   3378   HB  THR L 227     -36.166  22.580  28.848  1.00  0.00           H
ATOM   3379  HG1  THR L 227     -34.994  19.987  29.136  1.00  0.00           H
ATOM   3380 1HG2  THR L 227     -34.138  22.579  30.246  1.00  0.00           H
ATOM   3381 2HG2  THR L 227     -35.593  22.831  31.238  1.00  0.00           H
ATOM   3382 3HG2  THR L 227     -34.799  21.240  31.213  1.00  0.00           H
TER
# All scores below are weighted scores, not raw scores.
#BEGIN_POSE_ENERGIES_TABLE 3gbn_Ab_0005.pdb
label fa_atr fa_rep fa_sol fa_intra_rep fa_elec pro_close hbond_sr_bb hbond_lr_bb hbond_bb_sc hbond_sc dslf_fa13 rama omega fa_dun p_aa_pp yhh_planarity ref total
weights 1 0.55 0.9375 0.005 0.875 1.25 1.17 1.17 1.17 1.1 1.25 0.25 0.625 0.7 0.4 0.625 1 NA
pose -994.338 137.719 561.027 2.15688 -112.612 19.7634 -12.2069 -79.5391 -24.1449 -23.4441 -1.15166 -7.47192 71.8572 276.633 -29.8673 0.09431 13.9828 -201.541
GLU:NtermProteinFull_1 -1.58225 0.53996 1.45283 0.00353 0.06909 0 0 0 0 0 0 0.01109 6.53174 0 0 -1.96094 5.06505
VAL_2 -3.34255 0.45648 1.46378 0.01322 -0.08167 0 0 0 0 0 -0.16095 0.87346 0.30715 0.39992 0 0.97964 0.90848
GLN_3 -2.79445 0.10936 1.74929 0.00451 -0.52743 0 0 0 -0.35772 0 0 -0.09682 0.35321 2.59775 0.02034 0 -1.51717 -0.45911
LEU_4 -5.13483 0.73792 1.6574 0.00685 -0.16379 0 0 0 0 0 0.06265 0.2281 2.29891 -0.1217 0 0.76113 0.33264
VAL_5 -2.72905 0.12167 1.72074 0.00789 -0.45069 0 0 0 0 0 -0.27382 0.01969 0.02557 -0.49649 0 0.97964 -1.07485
```

How do I run a Rosetta command?

Toward high-resolution prediction and design of transmembrane helical protein structures

P. Barth, J. Schonbrun*, and D. Baker†

Department of Biochemistry and Howard Hughes Medical Institute, University of Washington, Seattle, WA 98195

```
$ROSETTA/main/source/bin/AbinitioRelax.linuxgccrelease -
database ../../rosetta_database -in:file:fasta ./input_files/
1elwA.fasta -in:file:native ./input_files/1elw.pdb -in:file:frag3 ./
input_files/aa1elwA03_05.200_v1_3 -in:file:frag9 ./input_files/
aa1elwA09_05.200_v1_3 -abinitio:relax -relax:fast -
abinitio::increase_cycles 10 -abinitio::rg_reweight 0.5 -
abinitio::rsd_wt_helix 0.5 -abinitio::rsd_wt_loop 0.5 -use_filters
true -psipred_ss2 ./input_files/1elwA.psipred_ss2 -kill_hairpins -
out:file:silent 1elwA_silent.out
-nstruct 10
```

OR

$ROSETTA/main/source/bin/AbinitioRelax.linuxgccrelease @options.txt

Use an options file for your runs

$ROSETTA/main/source/bin/AbinitioRelax.linuxgccrelease @options.txt

```
-in:file:
    -fasta ./input_files/1e1wA.fasta
    -native ./input_files/1e1w.pdb
    -frag3 ./input_files/aa1e1wA03_05.200_v1_3
    -frag9 ./input_files/aa1e1wA09_05.200_v1_3
-psipred_ss2 ./input_files/1e1wA.psipred_ss2
-abinitio:relax
-relax:fast
-abinitio::increase_cycles 10
-abinitio::rg_reweight 0.5
…
-out:file:silent ./output_files/1e1wA_silent.out
-nstruct 10
```

Other files: application-specific

- Span File: Defines which residues are in the membrane
- Loops File: Identifies the loop residues for loop closure
- Params File: Custom parameters for small molecules or non-canonical amino acids
- Constraint File: Experimentally derived restraints
- Fragment File: Short protein segments used for comparative modeling and de novo folding
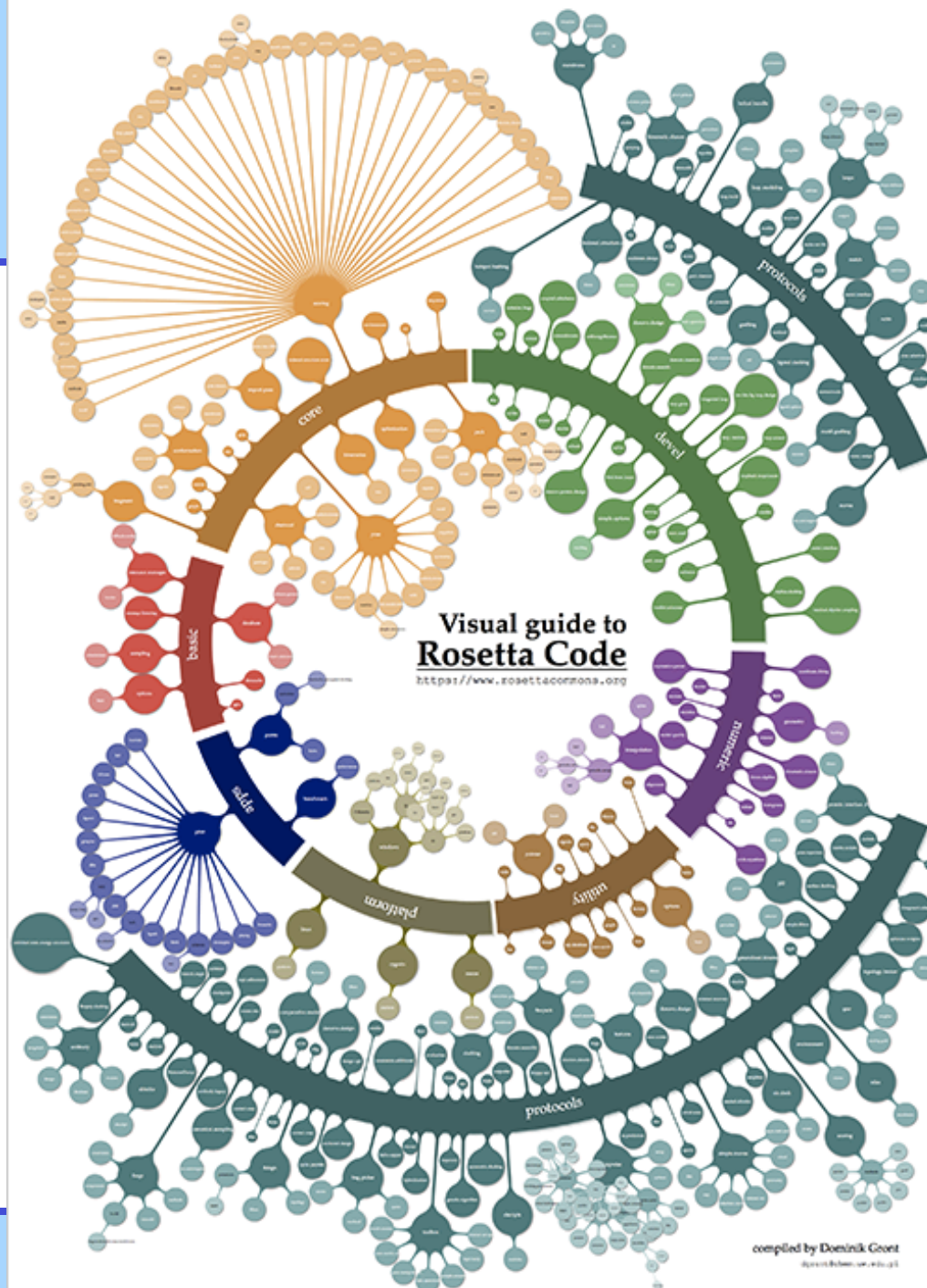- Res Files: Indicates which residue positions should be designed

Tracer output (log files)

```
core.init: Rosetta version exported  from http://www.rosettacommons.org
core.init: command: /dors/meilerlab/apps/rosetta/rosetta_2016.08.58479/main/source/bin/rosetta_scripts.default.linuxgccrelease @docking.options -parser:protocol docking
core.init: 'RNG device' seed mode, using '/dev/urandom', seed=1059677151 seed_offset=0 real_seed=1059677151
core.init.random: RandomGenerator:init: Normal mode, seed=1059677151 RG_type=mt19937
core.init: Resolved executable path: /dors/meilerlab/apps/rosetta/rosetta_2016.08.58479/main/source/build/src/release/linux/2.6/64/x86/gcc/5.2/default/rosetta_scripts.d
core.init: Looking for database based on location of executable: /dors/meilerlab/apps/rosetta/rosetta_2016.08.58479/main/database/
protocols.jd2.PDBJobInputter: Instantiate PDBJobInputter
protocols.jd2.PDBJobInputter: PDBJobInputter::fill_jobs
protocols.jd2.PDBJobInputter: pushed 3gbm_HA_3gbn_Ab.pdb nstruct indices 1 - 50
protocols.evaluation.ChiWellRmsdEvaluatorCreator: Evaluation Creator active ...
protocols.jd2.JobDistributor: Parser is present.  Input mover will be overwritten with whatever the parser creates.
protocols.jd2.PDBJobInputter: PDBJobInputter::pose_from_job
protocols.jd2.PDBJobInputter: filling pose from PDB 3gbm_HA_3gbn_Ab.pdb
core.chemical.ResidueTypeSet: Finished initializing fa_standard residue type set.  Created 384 residue types
core.chemical.ResidueTypeSet: Total time to initialize 0.43 seconds.
core.conformation.Conformation: Found disulfide between residues 7 461
...
protocols.rosetta_scripts.ParsedProtocol.REPORT: ============End report for =================
protocols.rosetta_scripts.ParsedProtocol.REPORT: ============Begin report for =================
protocols.rosetta_scripts.ParsedProtocol.REPORT: ============End report for =================
protocols.jd2.JobDistributor: 3gbm_HA_3gbn_Ab_full_0050 reported success in 381 seconds
protocols.jd2.JobDistributor: no more batches to process...
protocols.jd2.JobDistributor: 50 jobs considered, 50 jobs attempted in 16297 seconds
~
~
~
```

Options to control tracer output:

- Silence certain tracers:

    `-mute core.chemical.ResidueTypeSet`

- Change verbosity level (Error/Warning/Info/Debug/Trace)

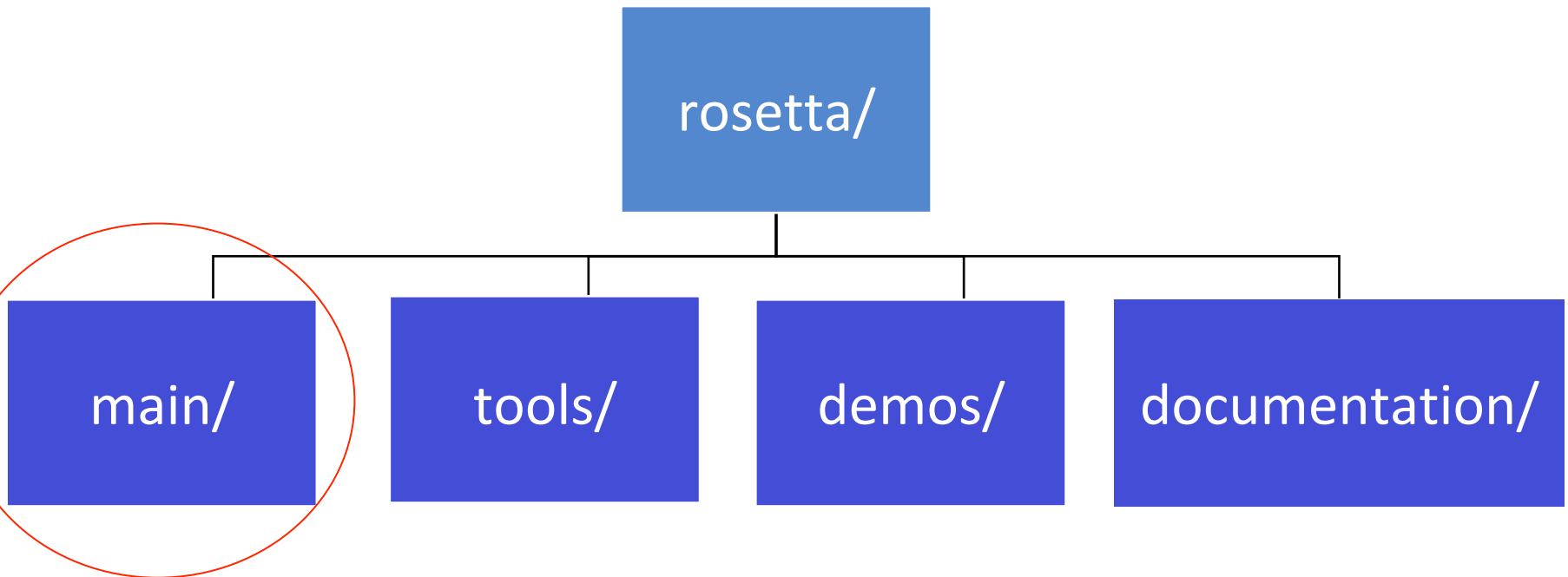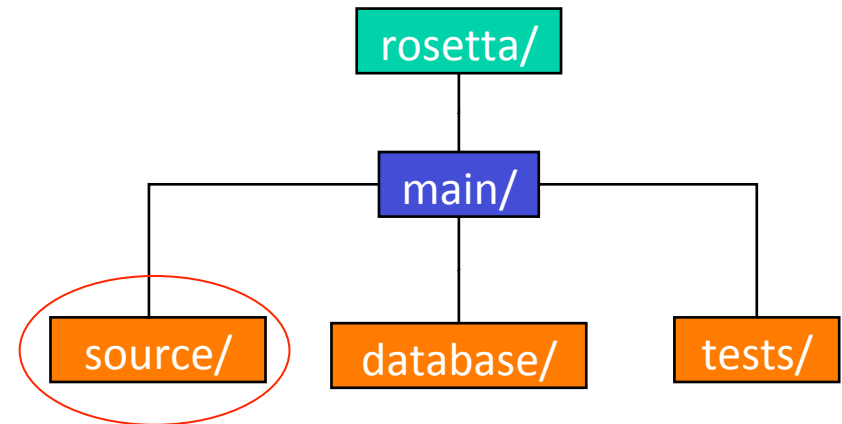   `-out:levels all:Warning core.init:Info`

Any Questions?

Visual guide to
Rosetta Code
https://www.rosettacommons.org

compiled by Dominik Gront

Basic Rosetta Structure

cd ~/rosetta_workshop/rosetta/

- Rosetta/main/source/bin/
  - Most applications you will run are calling programs within the bin directory
    - *rosetta_scripts.default.linuxgccrelease
    - *score_jd2.default.linuxgccrelease
    - relax.default.linuxgccrelease

- Rosetta/main/source/scons.py
  - Used for compiling
- Rosetta/main/source/src/
  - This is where all of the code lives
- Rosetta/main/source/scripts/
  - Some useful scripts live here (e.g. params file generation)

rosetta/
main/
source/   database/   tests/

The Rosetta database contains pre-defined information that an application needs for runs—users generally don't change things here.

Note: If you've compiled regularly, applications know where to search for things in the database without you telling it where to search.
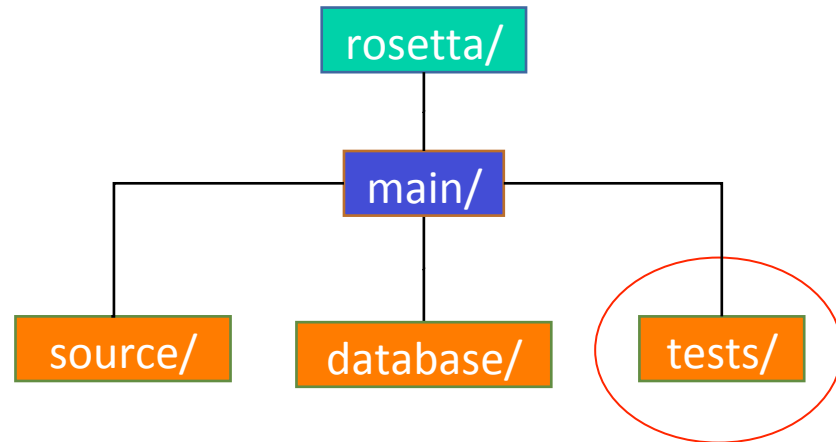
- Rosetta/main/database/scoring/
  - Default weights files
  - Rotamer libraries

- Rosetta/main/database/chemical/
  - Residue information--params files
  - Atom sets
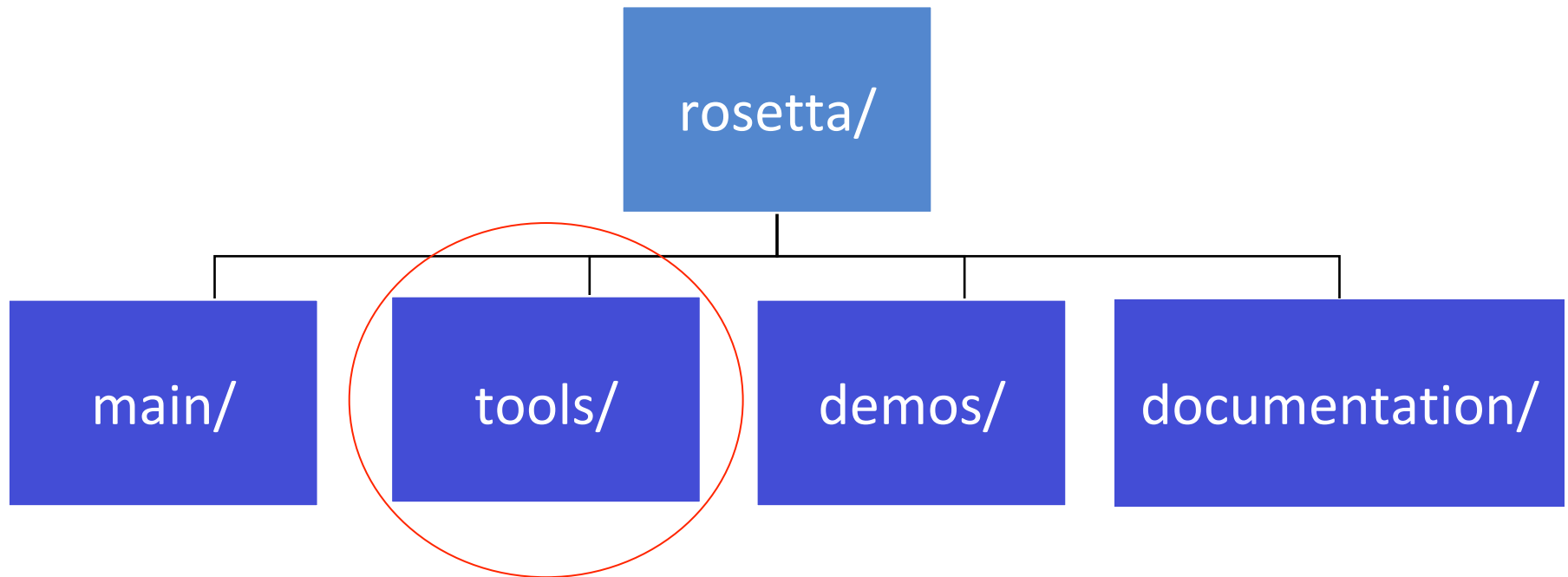
- Rosetta/main/tests/
  - This is where developers are developing and testing their new protocols

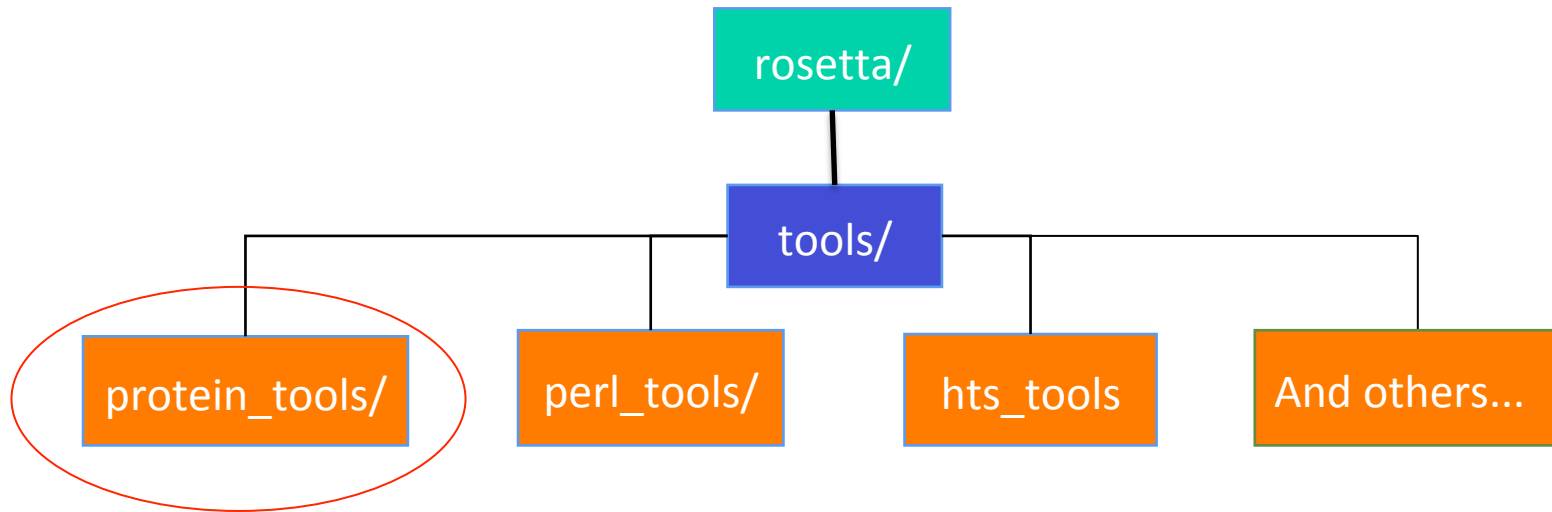# Basic Rosetta Structure

cd ~/rosetta_workshop/rosetta/

~/rosetta_workshop/rosetta/tools/

```
                        ┌─────────────┐
                        │   rosetta/  │
                        └──────┬──────┘
                               │
                        ┌──────┴──────┐
                        │    tools/   │
                        └──────┬──────┘
     ┌───────────────┬─────────┼─────────┬───────────────┐
┌──────────────┐ ┌──────────────┐ ┌──────────────┐ ┌──────────────┐
│ protein_tools/│ │  perl_tools/ │ │   hts_tools  │ │ And others...│
└──────────────┘ └──────────────┘ └──────────────┘ └──────────────┘
```

- These scripts are incredibly help for smaller, more basic tasks
- Used mainly to setup or analyze runs

Some useful scripts to be aware of:

clean_pdb.py*
- Makes a PDB "Rosetta-proof" and used at the beginning of almost any protocol

pdb_renumber.py
- Renumbering your PDB starting from 1

score_vs_rmsd.py
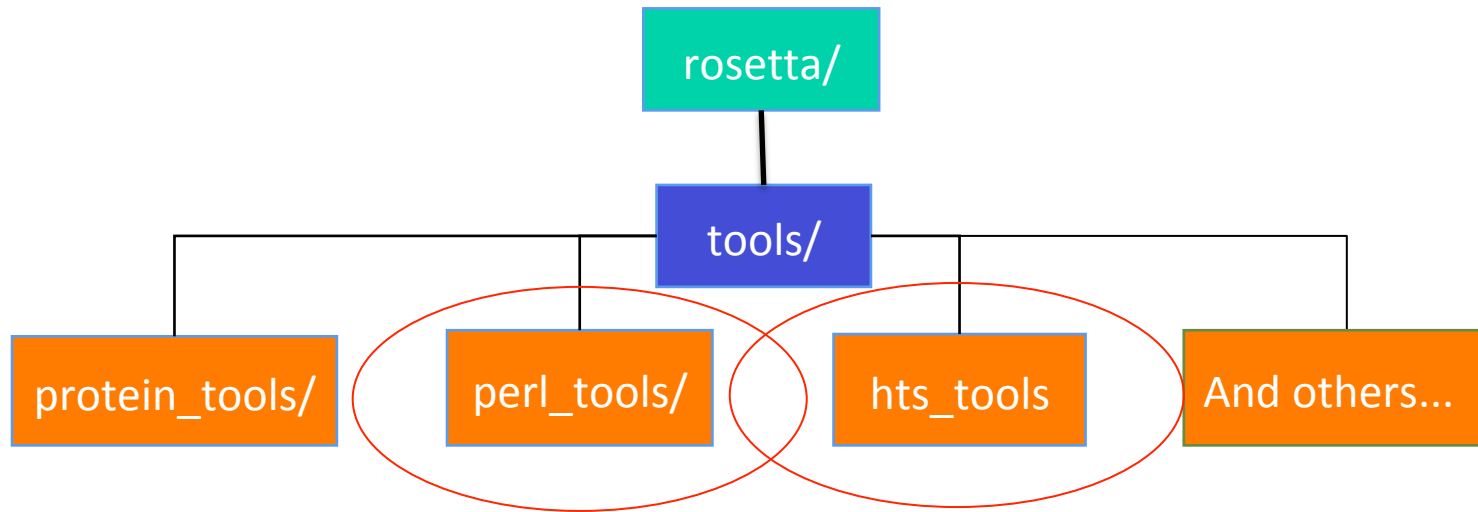- Setup for score vs. RMSD plots

top_n_percent.py
- Extracts tags (protein names) for top given percent of models based on score term

~/rosetta_workshop/rosetta/tools/perl_tools/
~/rosetta_workshop/rosetta/tools/hts_tools/

Some useful scripts to be aware of:

Rosetta/tools/perl_tools/
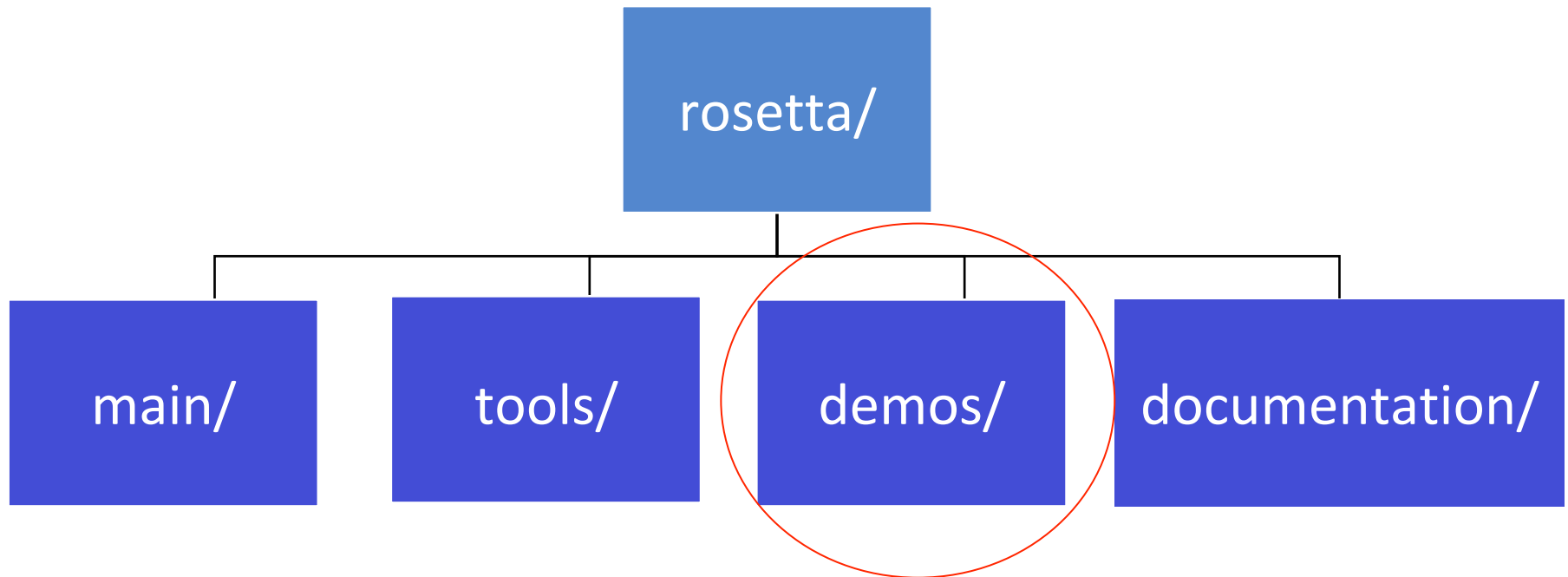- addchain.pl
- getFastaFromCoords.pl
- getCAcoords.pl

Rosetta/tools/hts_tools/
Scripts used mainly to setup high throughput screen runs (> 1000 compounds)

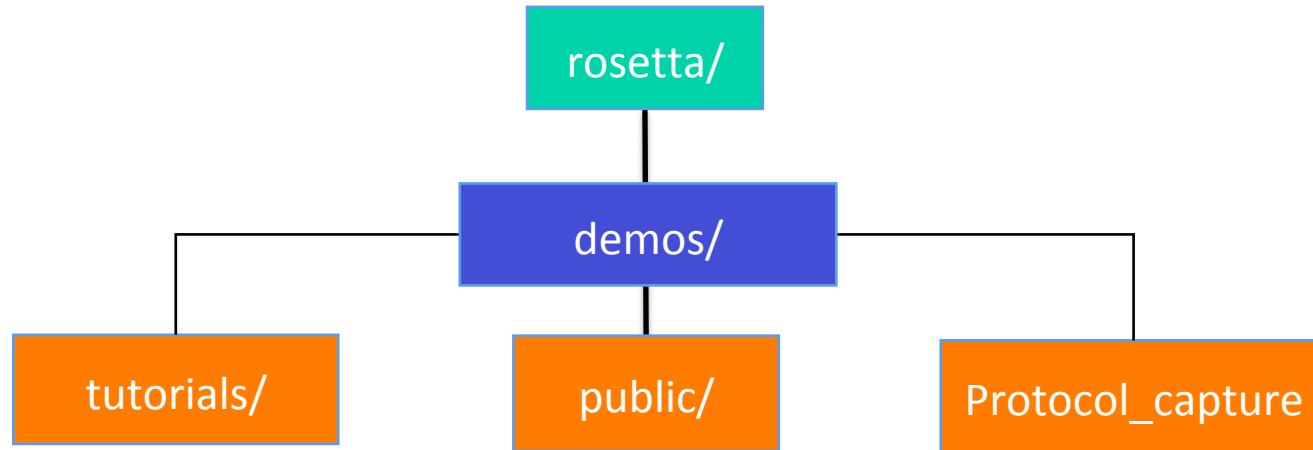# Basic Rosetta Structure

cd ~/rosetta_workshop/rosetta/

tutorials/
- Past tutorials
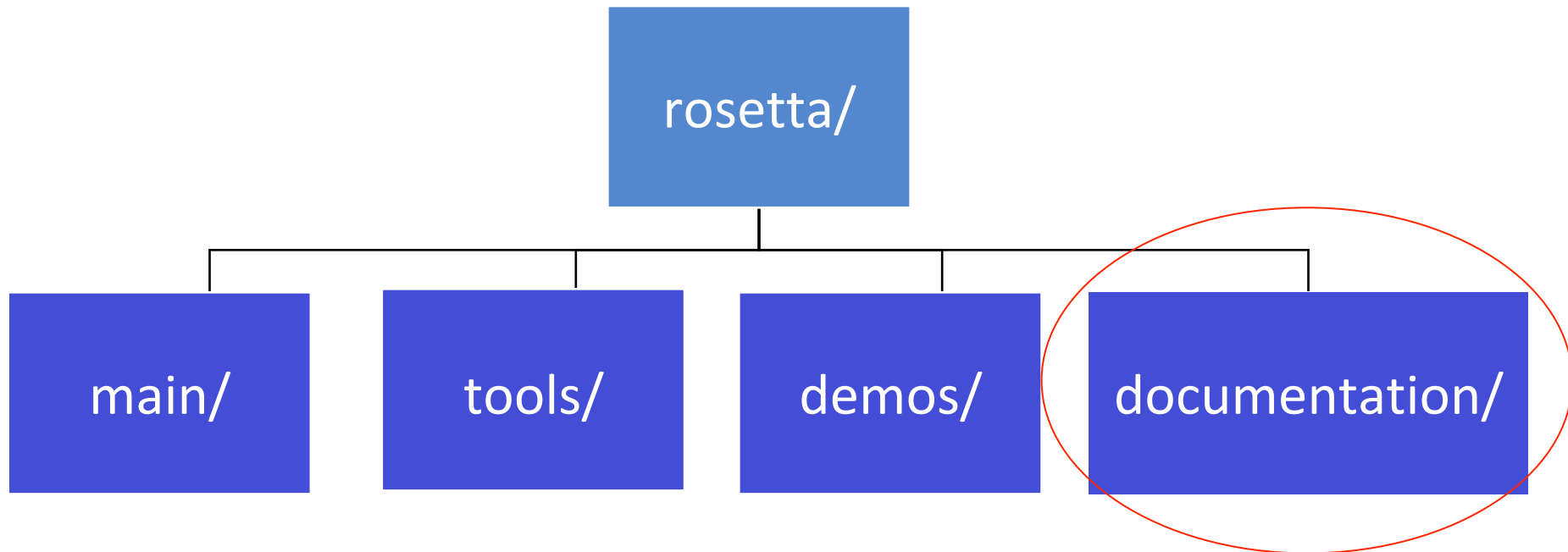
public/
- Protocol examples

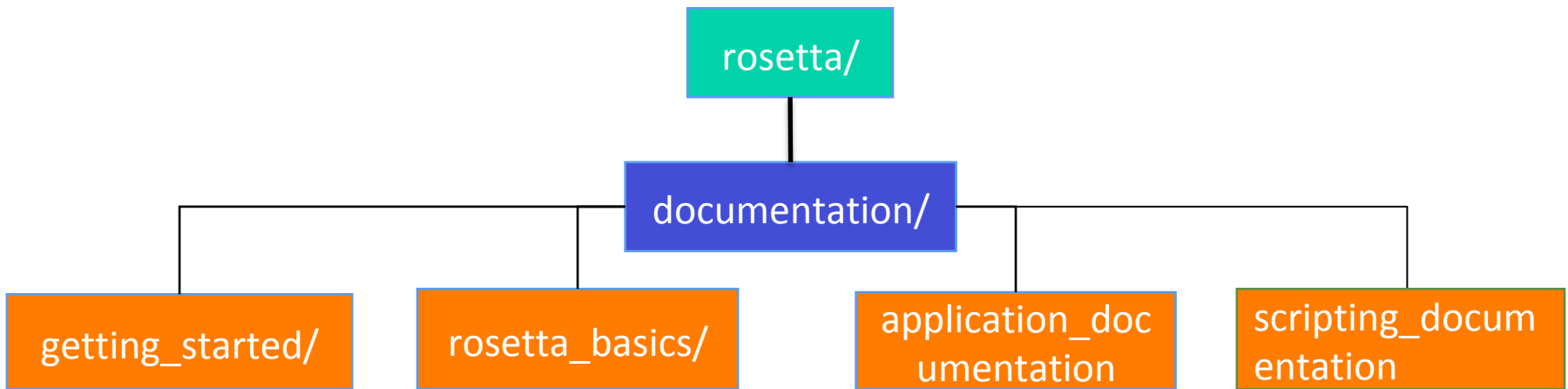protocol_capture/
- protocols associated with a publication

DISCLAIMER: May be out of date, always check Wiki/RosettaCommons/forum for latest information!

Basic Rosetta Structure

cd ~/rosetta_workshop/rosetta/

```
                        rosetta/
                           |
                    documentation/
     _____|_____
    |              |              |              |
getting_started/  rosetta_basics/  application_doc  scripting_docum
                                   umentation       entation
```

Very useful to go through when you're just getting started in
Rosetta or any structural biology software

Understanding general Rosetta concepts
  • Where to find FAQs (How long does
    this run take?)
  • Options list, file types

Protocols you can use
    General structural biology FAQs
  • How do I do X?

Other Rosetta Resources:

- https://www.rosettacommons.org/docs/latest/Home
  - Forum
  - Documentation
  - User guides
  - Everything Rosetta-related you could ever dream of!

GO HAVE FUN!